## BIOL4536, Fall 2023

## Exam#2 (PRACTICE PROBLEMS)

**Question 1.** True or False. BLAST always returns the optimal (highest scoring) local alignment of a query sequence against a database of sequences.

**Question 2.** True or False. RNA-Seq reads can align ambiguously to multiple places in the genome but DNA-Seq reads cannot.

**Question 3.** In BLAST, why do we need the expected score to be negative? Choose the one correct answer.

- (A) Because otherwise the score of random alignments increases as the sequences get longer.
- (B) So that the optimal aignment score could be negative.
- (C) So that the BLAST random walk eventually converges to zero.
- (D) This is a trick question and we do not actually need the expected score to be negative.

Question 4. (Choose the one correct answer) The null hypothesis for a BLAST *p*-values is

- (A) The database consists of randomly generated sequences.
- (B) The expected score of any alignment between the query and a database sequence is zero.
- (C) The query sequence is generated according to the background probabilities.
- (D) The query sequence is not related to any sequence in the database.

Question 5. (Choose the one correct answer) The BLAST index we discussed in class:

- (A) Maps words to their exact occurrences in sequences in the database.
- (B) Maps words to alignments to words of the same length in sequences in the database whose alignment scores exceed a fixed threshold.
- (C) Maps sequences in the database to their alignment scores with the query.
- (D) Maps sequences in the database to their reverse complements.
- (E) Maps words in the query sequence to the set of all possible words of the same length that they align to with scores exceeding a fixed threshold.

**Question 6.** Explain how a SAM file bitflag encodes multiple pieces of binary information into one number.

Question 7. (Choose the one correct answer) The FPKM normalization normalizes for:

- (A) Depth of sequencing and read length.
- (B) Depth of sequencing and gene length.
- (C) Gene length and read length.
- (D) Number of replicates and number of conditions.
- (E) Length of the genome and depth of sequencing.

Question 8. Which things on the left go with which things on the right? Draw lines to connect them.



**Question 9.** Consider the following coverage plot and junctions track. Which of the four isoforms is expressed? Circle all that apply.



**Question 10.** What is size of the maximal excursion in the following BLAST random walk that has a score of +1 for a match and -1 for a mismatch:



**Question 11.** Why do we perform polyA selection in RNA-Seq? And why does polyA selection cause a 3' bias in the signal across the transcripts?

**Question 12.** Suppose we are using RNA-Seq to perform a differential expression analysis. Suppose there are exactly 30, 100 genes and exactly 100 of them are differentially expressed. Suppose we use a

*p*-value cutoff of 0.01 to call a gene significant and suppose all 100 of the truly differential genes have *p*-value below this cutoff. Calculate what we expect the proportion of false-positives is in the set of all significant genes.

Answer:

**Question 13.** Consider an alignment of a read to the genome with the following start location and CIGAR string.

chr9 1025050 75M1000N25M

Does the read align to the following genome location?

chr9:1025201

Answer:

Question 14. In a fastQ record, like the one shown below, what does the fourth line represent?

Answer:

**Question 15.** The following is a coverage plot and junctions track of RNA-Seq data for one gene. The gene annotation is not shown. Explain why there must be at least two isoforms expressed.



**Question 16.** Refer to the same graphic as in Question 15. Assume there are exactly two expressed isoforms. Which of the following is true? Choose the one correct answer.

(A) Both isoforms are expressed at the same level.

- (B) The shorter isoform is expressed roughly 40 times higher than the longer one.
- (C) The longer isoform is expressed roughly 40 times higher than the shorter one.
- (D) It cannot be determined which is higher, even approximately.

**Question 17.** Give one reason why, when performing RNA-Seq, there can be real signal visible across the introns of genes.

**Question 18.** How many sequences are in the motif described by this sequence frequency logo? Write down one of them that is neither the most nor the least likely.



**Question 19.** Suppose the raw count for a gene in a sample is 30. Suppose there are 12,000,000 readpairs and the gene has length 4,000 bases. What is the FPKM normalized value?

Question 20. True or False. An RNA-Seq read can span multiple exons.

**Question 21.** True or False. You can have an RNA-Seq read such that no part of it aligns to any exons of any gene.

**Question 22.** True or False. In the quality string for a read in a fastQ file, a capital letter indicates lower quality than a punctuation mark.

**Question 23.** Suppose you have a fastQ file of data that you want to Bowtie align it to a specific chromosome of a particular species for which you have the genome sequence. What do you have to do first before you can perform the actual alignment step?

Question 24. Consider the alignment of a 100 base read given by this CIGAR string:

chr19 11554011 31M260N69M

One exon of the ELOF1 gene is shown below. True or False. The alignment given by the CIGAR string is consistent with having come from the mature (introns removed) ELOF1 gene? Note, the exon has length exactly 71 bases.

Scale 20 bases | 11,554,015 | 11,554,020 | 11,554,025 | 11,554,035 | 11,554,040 | 11,554,045 | 11,554,055 | 11,554,055 | 11,554,060 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 | 11,554,075 |

**Question 25.** True or False. If you zoom in enough on a coverage plot track in the UCSC Genome Browser then it displays the individual reads.

Question 26. In a ChIP-Seq assay coverage plot, the interesting locations are:

- (A) Where the valleys are.
- (B) Where the peaks transition into the valleys, and vice versa.
- (C) Where the peaks are.
- (D) It depends, it's where the peaks are in TF binding ChIP-Seq and where the valleys are in histone-modification ChIP-Seq.

Question 27. What does the following clustering of RNA-Seq samples indicate?



**Question 28.** Suppose we're testing 10,000 genes for differential expression and only ten of them are actually differentially expressed. How small does the *p*-value cutoff for significance have to be in order for us to expect that at least half of the significant genes are true positives? *Note:* "significant" here means its *p*-value is below the cutoff.

Question 29. In the histone mark H3K79me2 what does the "me2" mean?

Question 30. Explain the FPKM normalization, why we do it and how we do it?

**Question 31.** True or False. It is not possible to perform an RNA-Seq assay to sequence mRNA and short RNA ( $\leq$  50 bases) at the same time.

**Question 32.** True or False. PolyA selection in RNA-Seq retains coding RNA (mRNA) and discards everything else.

**Question 33.** True or False. For DNA-Seq we have to fragment the molecules because chromosomes are so long; while for RNA-Seq fragmentation is not necessary because molecules are so short.

**Question 34.** True or False. The gapped alignments that BLAST reports in its output are generated by Smith-Waterman.

Question 35. The "two-hit" improvement to BLAST was done to make it:

- (A) have more accurate *p*-values.
- (B) be less greedy.
- (C) run faster and use less resources.
- (D) have negative expected score.

Question 36. True or False. You need a different BLAST index for each substitution matrix.

**Question 37.** Shown below are coverage plots of two different genes from *two different* RNA-Seq assays. Give one reason the gene on the left might not actually be expressed higher than the one on the right.



**Question 38.** When making a coverage plot from the data depicted below. What is the maximum depth of coverage?



**Question 39.** What is the most frequent motif (or sequence) for a transcription factor with the following sequence frequency logo?



**Question 40.** True or False. In high-throughput sequencing, generating paired-end data allows us to sequence the fragments in their entirety.

**Question 41.** This genome browser graphic shows reads aligned to the gene FKBP14. Which the following are true? Circle all that apply.

(A) Both of these reads could align to the top isoform.

Answer:

(B) Both of these reads could align to the bottom isoform.

- (C) The top read could align to the top isoform while the bottom read could align the bottom isoform.
- (D) The top read could align to the botton isoform while the bottom read could align the top isoform.



**Question 42.** This is a protein BLAST random walk. What's the score in the substitution matrix for the A/D substitution?



Question 43. True or False. BLAST only reports ungapped alignments (no indels).

Question 44. True or False. BLAST works with any substitution matrix.

**Question 45.** Suppose we have an ungapped alignment between two DNA sequences. Suppose the score for any match is +1 and any mismatch is -1. Consider the BLAST random walk for this alignment. Suppose the max excursion is 12. True or False, the max excursion can occur at position 10 in the alignment.

**Question 46.** Suppose in a random walk heads are one step up and tails are one step down. Steps of the random walk are represented by the horizontal axis and cumulated step is represented on the vertical axis. Starting at the origin, draw in the random walk determined by the sequence of coin flips. Also identify the ladder points and the size of the max excursion.

H, H, T, H, H, T, T, T, T, H, H, T, T, T, H



**Question 47.** Why don't we consider an aligment with BLAST *p*-value of 0.01 as significant. Pick the one correct answer.

- (A) Because of all of the simplifying assumptions used to model alignment.
- (B) Because of multiple testing.
- (C) Because we never consider a *p*-value of 0.01 significant in any situtation.
- (D) It's not true, we do consider a BLAST *p*-value of 0.01 as significant.

Question 48. What does it mean that a histone modification is involved in "activation"?

Question 49. What are the two applications of ChIP-Seq that we discussed in class?

Answer:

Question 50. In the following BLAST alignment, what do the plus signs mean?

RecName: Full=H-2 class II histocompatibility antigen, A-D beta chain; Flags: Precursor [Mus musculus]

Sequence ID: P01921.1 Length: 265 Number of Matches: 1

See 1 more title(s) See all Identical Proteins(IPG)

Range 1: 41 to 227 GenPept Graphics

Vext Match A Previous Match

Score		Expect	Method	Identities	Positives	Gaps	
296 bits(759)		8e-102	Compositional matrix adjust.	140/187(75%)	161/187(86%)	1/187(0%)	
Query	1	ECYFTNG	TERVRLLTKYIYNREEYVRFDSDV( T+R+RL+T+YIYNREEYVR+DSDV(	GEYRAVTELGRPWA	EYWNPQKDEMDRV	RA 60 RA	
Sbjct	41	ECYYTNG	TQRIRLVTRYIYNREEYVRYDSDVO	GEYRAVTELGRPDA	EYWNSQPEILERT	RA 100	
Query	61	ELDTVCR E+DT CR	HNY-GLEELTTLQRRVEPTVTISPS	SRTEVLNHHNLLVC	SVTDFYPGQIKVR SVTDFYP +IKVR	WF 119 WF	
Sbjct	101	EVDTACR	HNYEGPETSTSLRRLEQPNVAISL	SRTEALNHHNTLVC	SVTDFYPAKIKVR	WF 160	
Query	120	RNDQEQT RN OE+T	AGVVSTPLIRNGDWTFQILVMLEM GV ST LIRNGDWTFO+LVMLEM	TPORGDVYTCHVEH	ASLQSPITVQWWP SL+SPITV+W	QS 179 OS	
Sbjct	161	RNGQEET	VGVSSTQLIRNGDWTFQVLVMLEM	TPHQGEVYTCHVEH	PSLKSPITVEWRA	QS 220	
Query	180	ESAQSKM ESA+SKM	186				
Sbjct	221	ESARSKM	227				

Answer:

**Question 51.** Suppose the CIGAR string for an alignment of a read to the genome is. Note: Recall that "D" means deletion, "I" means insertion and that is with respect to the genome (in other words deleted from or inserted in the genome). "S" means skip that many bases in the read.

## 25M475N32M2D4I3S

How long is the read?

Answer:

Question 52. Give one reason we might cluster RNA-Seq samples after quantifying.

Answer:

Question 53. Why is gene-level quantification easier than isoform level quantification?

**Question 54.** Suppose the genotype of a human at a SNP location is heterogenous A/G. Suppose we perform DNA-Seq and the read-depth at the SNP location is 5. Suppose each read has 50% probability of being maternal and 50% probability of being paternal. What is the probability that the reads all indicate the same nucleotide, thus leading to an erroneous (homozygous) genotype call at this location?

Answer:

**Question 55.** Assume the human genome is 3.6 billion nucleotides. Suppose we are performing 100-base single-end read DNA-Seq. What is the average depth-of-coverage if you have 360 million reads all

fully aligned?

Answer:

**Question 56.** Suppose a random walk is based on a coin flip, step +1 for a head and -1 for a tail. Suppose the coin is biased with 40% probability of a head. The long term behavior of the random walk is:

- (A) It will drift off to negative infinity.
- (B) It will converge to zero.
- (C) It will drift off to positive infinity.
- (D) None of the above.

**Question 57.** Consider the following coverage plot and junctions track. Is this data explained completely by the gene annotation that is displayed? Explain why or why not.



Question 58. Why do we deplete ribosomal RNA when we perform RNA-Seq?

**Question 59.** True or False. An RNA-Seq read can align to the genome in a location where there is no gene annotated.

**Question 60.** Describe what kind of search blastX is for. Answer:

**Question 61.** Suppose *X* is a discrete random variable that takes finitely many values x = 1, 2, ..., n and has probability function  $p_X(i)$ . Show using the formula for expected value that E[2X] = 2E[X].

**Question 62.** What is X - Y? *Hint: consider* d(b, c) + d(b, d) - d(a, c) - d(a, d).



Question 63. Why should we not use RNA-Seq for genotyping?